RESEARCH Open Access



# Addition of the Aβ42/40 ratio to the cerebrospinal fluid biomarker profile increases the predictive value for underlying Alzheimer's disease dementia in mild cognitive impairment

Inês Baldeiras<sup>1,2,3\*</sup>, Isabel Santana<sup>2,3,4</sup>, Maria João Leitão<sup>1,2</sup>, Helena Gens<sup>4</sup>, Rui Pascoal<sup>1</sup>, Miguel Tábuas-Pereira<sup>4</sup>, José Beato-Coelho<sup>4</sup>, Diana Duro<sup>4</sup>, Maria Rosário Almeida<sup>2</sup> and Catarina Resende Oliveira<sup>2,5</sup>

### **Abstract**

**Background:** Cerebrospinal fluid (CSF) biomarkers have been used to increase the evidence of underlying Alzheimer's disease (AD) pathology in mild cognitive impairment (MCI). However, CSF biomarker-based classification often results in conflicting profiles with controversial prognostic value. Normalization of the CSF A $\beta$ 42 concentration to the level of total amyloid beta (A $\beta$ ), using the A $\beta$ 42/40 ratio, has been shown to improve the distinction between AD and non-AD dementia. Therefore, we evaluated whether the A $\beta$ 42/40 ratio would improve MCI categorization and more accurately predict progression to AD.

**Methods:** Our baseline population consisted of 197 MCI patients, of which 144 had a follow-up  $\geq$  2 years, and comprised the longitudinal study group. To establish our own CSF A $\beta$ 42/40 ratio reference value, a group of 168 AD-dementia patients and 66 neurological controls was also included. CSF biomarker-based classification was operationalized according to the framework of the National Institute of Aging–Alzheimer Association criteria for MCI.

**Results:** When using the core CSF biomarkers (A $\beta$ 42, total Tau and phosphorylated Tau), 30% of the patients fell into the high-AD-likelihood (HL) group (both amyloid and neurodegeneration markers positive), 30% into the low-AD-likelihood group (all biomarkers negative), 28% into the suspected non-Alzheimer pathophysiology (SNAP) group (only neurodegeneration markers positive) and 12% into the isolated amyloid pathology group (only amyloid-positive). Replacing A $\beta$ 42 by the A $\beta$ 42/40 ratio resulted in a significant increase in the percentage of patients with amyloidosis (42–59%) and in the proportion of interpretable biological profiles (61–75%), due to a reduction by half in the number of SNAP cases and an increase in the proportion of the HL subgroup. Survival analysis showed that risk of progression to AD was highest in the HL group, and increased when the A $\beta$ 42/40 ratio, instead of A $\beta$ 42, combined with total Tau and phosphorylated Tau was used for biomarker-based categorization.

**Conclusions:** Our results confirm the usefulness of the CSF A $\beta$ 42/40 ratio in the interpretation of CSF biomarker profiles in MCI patients, by increasing the proportion of conclusive profiles and enhancing their predictive value for underlying AD.

Keywords: Mild cognitive impairment, CSF biomarkers, Aβ42/40 ratio, Alzheimer's disease

Full list of author information is available at the end of the article



<sup>\*</sup> Correspondence: ines.baldeiras@sapo.pt

<sup>&</sup>lt;sup>1</sup>Laboratory of Neurochemistry, Neurology Department, Centro Hospitalar e Universitário de Coimbra, 3000-075 Coimbra, Portugal

<sup>&</sup>lt;sup>2</sup>Center for Neuroscience and Cell Biology, University of Coimbra, 3004-504 Coimbra, Portugal

### **Background**

Alzheimer's disease (AD) is the leading cause of dementia worldwide and the most common neurodegenerative disease, affecting 4.6–8.7% of people over the age of 60 [1]. The pathophysiological process of AD is thought to begin many years before its clinical diagnosis [2, 3] and is generally hypothesized to be initiated by abnormal amyloid processing, followed by neuronal dysfunction and structural brain changes, which ultimately lead to cognitive impairment and dementia [4]. The interest in capturing the earliest stages of AD has been supported by the development of biomarkers of the disease like the core cerebrospinal fluid (CSF) AD biomarkers, amyloid positron emission tomography (PET) imaging and evidence of hippocampal atrophy on MRI. These biomarkers, reflecting both amyloid deposition and neuronal injury, have been incorporated into new diagnostic criteria, like those proposed by the National Institute of Aging-Alzheimer Association (NIA-AA) for AD dementia [5], mild cognitive impairment (MCI) [6] or preclinical states [7].

Core CSF biomarkers for AD are Aβ42, which is found in low concentrations in AD, probably reflecting brain amyloid deposition, total tau (t-Tau) at high concentrations representing cortical neuronal loss and phosphorylated tau (p-Tau) also at high concentrations, reflecting cortical tangle formation [8]. These markers have shown high diagnostic accuracy for established AD [9], and they may also be used to identify AD before onset of dementia at the stage of MCI, as shown in both single-center studies [10, 11] and large-scale heterogeneous multicenter studies [12-14]. However, low specificity regarding the distinction between AD and other types of degenerative dementias is still an issue and concern remains regarding the inter-laboratory variability of these CSF biomarkers and the lack of harmonization between centers [15]. In fact, several international standardization initiatives have already been launched to address these standardization issues [16, 17], and major advancements have been made in the field [18].

Recently, in order to improve the accuracy for AD diagnosis, other CSF biomarkers related to amyloid-beta (A $\beta$ ) metabolism have been studied [19]. A $\beta$  is produced by the sequential proteolytic cleavage of amyloid precursor protein (APP) by  $\beta$ -secretases and  $\gamma$ -secretases [20], resulting in at least five different C-terminally truncated A $\beta$  isoforms. The most abundant A $\beta$  isoform in CSF is A $\beta$ 40 [19] which is less prone to aggregation, and thus, at least theoretically, a more direct measure of total brain A $\beta$  content. It is thus conceivable that A $\beta$ 42 concentration, the A $\beta$  isoform with a higher aggregation tendency, depends not only on the physiologic status (presence or absence of amyloid aggregates) but also on the total amount of A $\beta$  peptides in the CSF, reflecting

different efficiency of APP processing. Therefore, the use of the Aβ42/40 ratio is thought to more accurately reflect the changes in Aβ metabolism in AD than Aβ42 alone, as it corrects for individual baseline differences in both high and low amyloid-producing individuals [21]. Indeed, results from several groups demonstrated that normalization of the CSF AB42 concentration to the level of total Aβ peptides, using the Aβ42/40 ratio, improved the distinction between dementia and controls [22, 23] and also between AD and non-AD dementia [24–26], particularly in cases with ambiguous core CSF biomarker profiles (i.e., isolated reduction of Aβ42 or elevation of t-Tau/p-Tau) [27-29]. Recently, the CSF Aβ42/40 ratio has also been shown to be superior to Aβ42 alone in reflecting amyloid PET status (positive vs negative) [26, 30]. The added value of the CSF A $\beta$ 42/40 ratio for predicting AD in patients with MCI has so far been less studied. Hansson et al. [31] followed 131 MCI patients for 4-6 years and showed that the Aβ42/40 ratio was better than the Aβ42 concentration in identifying incipient AD in MCI. On the contrary, Parnetti et al. [32], in a study employing 90 MCI patients followed for up to 4 years, stated that the performance of the Aβ42/ 40 ratio was not superior to A $\beta$ 42 alone and that the Aβ42/p-tau ratio was the best parameter to predict conversion to AD in MCI patients. Also, in a smaller study, Brys et al. [33] had already shown that the A $\beta$ 42/40 ratio was inferior to p-Tau alone in predicting decline from MCI to AD. Very recently, a multicenter memory clinic cohort study from the German Dementia Competence Network [34], with 115 MCI patients, also indicated that Aβ42/40 ratio was not consistently superior to Aβ42 alone for predicting short-time progression to AD.

The NIA-AA guidelines for MCI due to AD propose categorizing MCI according to the individual likelihood of underlying AD pathophysiology, according to their biomarker profile [6]. In these guidelines, the highest likelihood category is characterized by biomarker findings pointing to the presence of AD pathophysiology, whereas the lowest likelihood category is characterized by findings not typical for AD. This categorization also includes subgroups of conflicting biomarker results, namely patients with biomarkers positive for amyloidosis but negative for neurodegeneration and patients with normal amyloid markers but positive for neurodegeneration. A number of studies, using CSF Aβ42, t-Tau and p-Tau, as well as imaging markers, have investigated the prognostic relevance of these biomarker-based categories in MCI patients [35-42]. General agreement exists as to the risk of progression to AD being higher in patients with all biomarkers positive for AD and lowest in patients with no positive biomarkers for AD. However, the biological significance and the prognosis of patients who

fall into conflicting biomarker categories are still controversial.

Considering the data described, showing the relevance of A $\beta$ 40 inclusion in the panel of CSF biomarkers as a way of reducing diagnostic uncertainty, we hypothesize that using the A $\beta$ 42/40 ratio would significantly improve MCI categorization according to the NIA-AA guidelines by reducing the number of patients with conflicting biomarker results. Therefore, in this study, we propose to: first, using a group of AD patients and neurological controls, establish a cutoff value for the CSF A $\beta$ 42/40 ratio in our population; second, in an amnestic MCI cohort, assess the changes in MCI patients' NIA-AA criteria classification induced by the inclusion of the A $\beta$ 42/40 ratio in their CSF biomarker profile; and, third, evaluate the prognostic value of this new classification for AD-type dementia at follow-up.

### Methods

### **Subjects**

This study included 431 subjects (168 AD-dementia patients, 197 MCI patients and 66 neurological controls) for whom CSF-AD biomarker assessment, including A $\beta$ 40, was available.

AD-dementia and MCI patients were recruited at the Dementia Clinic, Neurology Department of Coimbra University Hospital, Coimbra, Portugal. The baseline study and follow-up protocol have already been published elsewhere [43]. Patients were enrolled in a systematic way and had biannual clinical observation and annual neuropsychological and functional evaluations. All patients underwent a thorough biochemical, neurological and imaging (CT or MRI and SPECT) evaluation. PET and genetic studies were more restricted, although considered in younger patients. At baseline, a neurologist completed a medical history with the patient and the caregiver, and conducted a general physical, neurological and psychiatric examination as well as a comprehensive diagnostic battery protocol, including: cognitive instruments such as the Mini Mental State Examination (MMSE) [44] Portuguese version [45], The Montreal Cognitive Assessment (MoCA) [46] Portuguese version [47], the Alzheimer Disease Assessment Scale—-Cognitive (ADAS-Cog) [48, 49] Portuguese version [50] and a comprehensive neuropsychological battery with normative data for the Portuguese population (Lisbon Battery for Dementia Assessment (BLAD)) [51] exploring memory (Wechsler Memory Scale subtests) and other cognitive domains (including language, praxis, executive functions and visuoconstrutive tests); and standard staging scales which provide objective information about subject performance in various domains, including the Clinical Dementia Rating scale (CDR) [52] for global staging, the Disability Assessment for Dementia (DAD) [53, 54] for evaluation of functional status and the Neuropsychiatric Inventory (NPI) [55, 56] to characterize the psychopathological profile, including the presence of depression. All of the available information (baseline cognitive test, staging scales, clinical laboratory and imaging studies) was used to reach a consensus research diagnosis. A similar approach was used for follow-up annual evaluations.

MCI patients included in this study were of the amnestic type and the diagnosis was made in accordance with the criteria defined by Petersen et al. [57] and, more recently, the framework for MCI due to AD proposed by NIA-AA criteria [6]. Petersen et al.'s criteria were operationalized as follows: a subjective complaint of memory decline (reported by the subject or an informant); an objective memory impairment (considered when scores on standard Wechsler memory tests were > 1.5 SDs below age/education-adjusted norms) with or without deficits in other cognitive domains; normal general cognition suggested by normal scores for the MMSE and MoCA using the Portuguese cutoff scores [45, 58]; largely normal daily life activities, evaluated with a functional scale (DAD); and absence of dementia, indicated by a CDR rating of 0.5. All patients were in a stable condition, without acute comorbidities. As exclusion criteria for enrolment, we considered a significant underlying medical or neurological illness revealed by laboratory tests or imaging; a relevant psychiatric disease, including major depression, suggested in the medical interview and confirmed by the GDS; and CT or MRI demonstration of significant vascular burden [59] (large cortico-subcortical infarct; extensive subcortical white matter lesions superior to 25%; unilateral or bilateral thalamic lacunes; lacune in head of caudate nucleus; more than two lacunes).

MCI cases were followed up with this comprehensive protocol until they developed dementia, or until they had been cognitively stable for at least 2 years, comprise the longitudinal study group. This group was further dichotomized into those who were cognitively stable and those who developed dementia due to AD. Patients who developed types of dementia other than AD were further excluded from analysis. Conversion to AD required fulfilling clinical diagnostic criteria for probable AD (see later) and was confirmed by the coordinator of the clinical study. As these criteria are not fully operational and the conversion status decision has some uncertainty and subjectivity, patients in this study were classified as having undergone conversion based on: objective evidence, by cognitive testing, of decline to dementia using the MMSE, the MoCA and the ADAS-Cog scores and qualitative evaluation (i.e., impairment of memory plus another domain); and changes in global CDR rating from 0.5 to 1 or more, confirming the cognitive profile of dementia and loss of autonomy.

For the biomarker-based subject classification, we used the core CSF biomarkers for AD, operationalized according to the framework of the NIA-AA criteria for MCI and preclinical forms [6, 7, 60]. Subjects were classified into the low-AD-likelihood (LL) group if both amyloid (i.e., CSF A $\beta$ 42) and neuronal injury markers (i.e., CSF t-tau and p-tau) were normal, into the high-AD-likelihood (HL) group if both amyloid and at least one neuronal injury marker were abnormal, or into one of the two conflicting biomarker groups: the isolated amyloid pathology (IAP) group if the amyloid marker was abnormal and neuronal injury markers were normal, or the suspected non-Alzheimer pathophysiology (SNAP) group if at least one neuronal injury marker was abnormal and the amyloid marker was normal.

Dementia was diagnosed according to the *Diagnostic and Statistics Manual for Mental Disorders*—fourth edition text review (DSM-IV-TR) criteria [61], and AD according to the National Institute of Neurological and Communicative Disorders and Stroke–Alzheimer's Disease and Related Disorders (NINCDS-ADRDA) criteria [62] and, more recently, the 2011 NIA-AA criteria [5]. These cases were classified as probable AD dementia according to clinical and neuroimaging features.

In this study we also included 66 neurological controls. Most of these individuals suffered from acute or chronic headaches, and a lumbar puncture was performed as part of their routine diagnostic evaluation in order to exclude bleeding or inflammation; in some cases, this procedure was considered in the investigation of a peripheral polyneuropathy. In both situations, the CSF cytochemical evaluation was normal and a major CNS disease was excluded. In their brief cognitive evaluation they showed no subjective cognitive complaints, were independent in their instrumental daily life activities and most of them were still professionally active.

### Laboratory determinations

CSF samples were collected from patients and neurological controls as part of their routine clinical diagnosis investigation. Preanalytical and analytical procedures were done in accordance with previously proposed protocols [63]. Briefly, CSF samples were collected in sterile polypropylene tubes, immediately centrifuged at  $1800 \times g$  for 10 min at 4 °C, aliquoted into polypropylene tubes and stored at -80 °C until analysis.

CSF A $\beta$ 42, t-Tau and p-Tau were measured separately, in duplicate, by commercially available sandwich ELISA kits (Innotest; Innogenetics/Fujirebio, Ghent, Belgium), as described previously [22, 64]. These assays were performed sequentially in a clinical routine setting between 2010 and 2017, with mean intra-assay coefficients of variation (CVs) of 4.2% for A $\beta$ 42, 4.5% for t-Tau and 4.2% for p-Tau and inter-assay CVs of 8.1% for A $\beta$ 42,

7.0% for t-Tau and 7.2% for p-Tau. CSF A $\beta$ 40 was also measured by ELISA, using the recently validated kit also from Fujirebio [65], following the manufacturer's instructions. In our hands, the intra-assay CV of this method was 3.8  $\pm$  1.8% (mean  $\pm$  SD) and the inter-assay CV was 13.2  $\pm$  4.0%, and therefore very similar to what has been reported [65]. A $\beta$ 40 assays were also performed in duplicate between November 2016 and March 2017.

External quality control of the assays was performed under the scope of the Alzheimer's Association Quality Control Program for CSF Biomarkers [66]. In this study, we established cutoff values for core CSF-AD biomarkers for this particular population by employing receiver operating characteristics (ROC) curve analysis between AD-dementia patients and controls, as reported previously [64]. According to these cutoff values (A $\beta$ 42 = 585 pg/ml, t-Tau = 244 pg/ml, p-Tau = 38 pg/ml), core CSF-AD biomarkers were classified as normal/abnormal.

Blood samples were also collected from MCI and AD patients for Apolipoprotein E (APOE) genotyping. DNA was isolated from whole EDTA blood using a commercial kit (Roche Diagnostics GmbH, Manheim, Germany), as described by the manufacturer. The analysis of the two polymorphisms at codons 112 and 158 of the *APOE* gene (rs429358 and rs7412) was performed by PCR-RFLP assay, as described previously [67].

### Statistical analysis

Statistical analyses were performed using the Statistical Package for the Social Sciences (SPSS, version 20.0; IBM SPSS, Chicago, IL, USA). Normality of continuous variables was assessed by the Kolmogorov-Smirnov test. For normally distributed continuous variables, one-way ANOVA followed either by the Bonferroni post test (when variance was homogeneous between groups) or the Games-Howell post test (when variance was not homogeneous between groups) was performed to assess the statistical significance of the difference between means. When continuous variables did not show normal distribution, the Kruskal-Wallis test was used, followed by the Dunn-Bonferroni post-hoc test. For CSF biomarkers, age was entered as a covariate in the analysis. Group differences between categorical variables were examined using the  $\chi^2$  test with Yate's correction for small sample sizes (n < 30). McNemar's test for paired proportions was used for testing differences between the proportions of cases with amyloidosis and with conclusive/ambiguous CSF biomarker profiles. ROC curve analysis was used to evaluate the diagnostic accuracy of the CSF markers or their ratios between AD-dementia patients and controls, and also of the predicted probabilities derived from the logistic regression models used to identify the best predictors of conversion to AD. The ROC curve was also used to establish the optimal cutoff

value, by selecting the value that yielded the highest Youden index calculated as: sensitivity + (specificity - 1). The ROC curves were compared according to the AUC comparison method of Hanley and McNeil [68] using MedCalc (version 11.6; MedCalc Software, Mariakerke, Belgium). Binary logistic regression analysis (enter method) was used to identify predictive markers of conversion to AD, with conversion as the dependent variable and age, gender, education, follow-up time, APOE genotype, baseline MMSE, CSF Aβ42, Aβ40, t-Tau and p-Tau levels as independent variables. Variables with a regression coefficient significantly different from 0 (associated p < 0.05) were considered to be contributing significantly to the prediction of the outcome variable. Survival analysis was used to assess the probability of conversion to AD in the different MCI subgroups. Kaplan-Meier survival curves were plotted and the survival distributions of the different MCI subgroups were compared by the logrank test. Survival time was calculated as the interval from the initial baseline evaluation to the diagnosis of dementia. For patients who remained nondemented, the survival time was censored at the date of the last clinical assessment. Cox proportional hazards models, corrected for age, gender, education, follow-up time, ApoE genotype and baseline MMSE score, were used to test the predictive ability for AD-type dementia of the different MCI groups.

### Results

### Characteristics of the study population

Demographic, clinical, genetic and biomarker data of the baseline study population are presented in Table 1. No differences in gender distribution were seen between groups, but the control group was significantly younger than the patient groups, and therefore age was entered as a covariate in the cognitive and CSF marker comparisons. Similar age of onset was seen between AD-dementia and MCI patients, but, as expected, AD-dementia patients presented with a more severe cognitive impairment (significantly lower MMSE and MoCA and higher ADAS-Cog scores) than MCI patients at baseline. The percentage of APOE-ε4 carriers in AD-dementia and MCI patients was over 40%, considerably higher than we showed previously in a Portuguese control population [69]. As reported previously [22, 64], CSF Aβ42 was lower and t-Tau and p-Tau were higher in AD-dementia patients compared to controls, while MCI patients had intermediate and significantly different values in relation to the two other groups. A significant increase in the t-Tau/Aβ42 ratio and a decrease in the Aβ42/p-Tau ratio in AD-dementia patients compared to controls was also observed, while MCI patients had intermediate and significantly different values from the other groups. CSF Aβ40 was similar between AD-dementia and controls, but was significantly higher in MCI patients in relation to AD-dementia patients only. This resulted in a decreased A $\beta$ 42/40 ratio in AD and MCI patients in relation to controls, and also in AD-dementia patients compared with the total MCI group.

### Establishment of a cutoff value for the CSF Aβ42/40 ratio

By comparing the A $\beta$ 42/40 ratio between AD-dementia patients and controls, as recommended by the STARD criteria [70], a cutoff value of 0.068 (with lower values indicative of AD) was established. This ratio had a sensitivity of 79% and a specificity of 86% to distinguish between AD-dementia patients and controls, with an AUC of 0.874 (95% CI 0.827–0.921). These accuracy parameters are similar to what we have reported previously for A $\beta$ 42 alone in a larger AD cohort [64] and are no different from that presented by A $\beta$ 42 alone in this particular AD-dementia population (sensitivity = 82%, specificity = 83%, AUC = 0.882, 95% CI 0.837–0.927, p = 0.748) (Additional file 1: Figure S1).

# Characterization of MCI biomarker-based subgroups according to the core CSF biomarkers

The MCI group was then classified into MCI subtypes according to their core CSF biomarkers (Aβ42, t-Tau and p-Tau): LL, 59 (29.9%); HL, 62 (31.5%); IAP, 21 (10.6%); and SNAP, 55 (27.9%). Noteworthy, the percentage of MCI patients with injury markers (59.3%) was higher than those with amyloidosis (42.1%). Table 2 presents the demographic, clinical, genetic and biomarker data of these subgroups. There were no significant differences regarding gender, years of education and time of follow-up, but the HL and SNAP patients were older at baseline and at onset of the symptoms (p = 0.001 vs LL group). Regarding the cognitive tests, both the MMSE and the MoCA mean scores were significantly lower in the HL group in comparison with the LL group (p < 0.001 and p = 0.004, respectively). Conversely, the ADAS-Cog mean score was higher both in the HL and SNAP groups vs the LL group (p = 0.005 and p = 0.015, respectively), again indicating greater cognitive impairment. As expected, differences were significant in terms of Aβ42 levels between HL and IAP vs SNAP and LL, as well as in terms of neuronal injury markers between SNAP and HL vs IAP and LL (p < 0.001 for all comparisons). Regarding Aβ40, a significant reduction was seen in the IAP group (p = 0.032 in relation to the HL group), while a significant increase was observed in the SNAP group in comparison with all the other groups (p < p)0.001 for all comparisons). Interestingly, in cases with an ambiguous biological profile (IAP + SNAP), Aβ40 levels were significantly more dispersed (ranged between 2899 and 41,282 pg/ml) than in patients with conclusive core CSF biomarkers (LL + HL; ranged between 3516 and 28,908 pg/ml, p = 0.001). A significantly increased Aβ42/40 ratio was seen in the LL group (p < 0.001 for

**Table 1** Demographic, clinical, genetic and biomarker data of the study population

	Controls ( $n = 66$ )	AD-dementia( $n = 168$ )	MCI total( $n = 197$ )	MCI-St(n = 74)	MCI-AD(n = 70)
Gender (M/F)	27/39	56/112	68/127	25/49	26/44
Age (years)	58.8 ± 11.9	68.1 ± 8.8***	67.1 ± 9.4***	65.2 ± 9.3*	71.0 ± 8.0***,§§
Age onset (years)		64.6 ± 8.7	64.2 ± 9.3	60.5 ± 9.4	68.8 ± 7.7 <sup>Y,§§§</sup>
Education (years)		$5.5 \pm 4.0$	$6.0 \pm 3.8$	$5.7 \pm 3.6$	$5.8 \pm 3.9$
MMSE	28.4±1.8	17.2 ± 5.9***	$25.9 \pm 4.0***,777$	$26.5 \pm 4.9***,777$	$25.1 \pm 3.3***,\gamma\gamma\gamma,\S$
MoCA		10.5 ± 5.1	$17.7 \pm 5.6^{YYY}$	$19.9 \pm 4.9^{YYY}$	$15.5 \pm 5.3^{\text{YY},\$\$\$}$
ADAS-Cog		26.0 ± 12.3	$11.8 \pm 6.1^{YYY}$	$9.0 \pm 4.3^{YYY}$	$14.7 \pm 6.3^{YYY,§§§}$
APOΕ-ε4 (%)		46%	42%	28%	60% <sup>§§§</sup>
Aβ42 (pg/ml)	928 ± 435	455 ± 225***	689 ± 322***,YYY	785 ± 309*, YYY	476 ± 180***,§§§
Aβ40 (pg/ml)	10,055 ± 4314	9229 ± 3668	11,097 ± 4635 <sup>YYY</sup>	$10,916 \pm 4379^{YY}$	10,157 ± 3381
Aβ42/40 ratio	$0.100 \pm 0.034$	0.055 ± 0.029***	$0.070 \pm 0.039^{***,YYY}$	$0.083 \pm 0.046^{*,YYY}$	$0.050 \pm 0.020^{***,§§§}$
t-Tau (pg/ml)	196 ± 95	543 ± 363***	$374 \pm 269***,YYY$	$253 \pm 151^{YYY}$	511 ± 307***,§§§
p-Tau (pg/ml)	33 ± 15	64 ± 37***	50 ± 28**, YYY	$38 \pm 19^{YYY}$	62 ± 30***,§§§
t-Tau/Aβ42	$0.22 \pm 0.11$	1.53 ± 1.36***	$0.74 \pm 0.73^{***,YYY}$	$0.41 \pm 0.39^{YYY}$	$1.18 \pm 0.85^{***,§§§}$
Aβ42/p-Tau	32.0 ± 15.3	9.3 ± 8.2***	18.6 ± 14.2***,YYY	$25.7 \pm 16.3^{YYY}$	9.7 ± 6.6***,§§§

Data expressed as mean ± standard deviation, except for APOE expressed as percentage of \$\xi\$4 carriers

MMSE and MoCA, higher scores correspond to better performance; ADAS-Cog, lower scores correspond to better performance. For MMSE and CSF biomarkers, data were adjusted for age

AD Alzheimer's disease, MCI mild cognitive impairment, MCI-St stable mild cognitive impairment, MCI-AD mild cognitive impairment patients who progress to Alzheimer's disease, M male, F female, MMSE Mini Mental State Examination, MoCA Montreal Cognitive Assessment, ADAS-Cog Alzheimer Disease Assessment Scale—Cognitive, APOE Apolipoprotein E, Aβ42 42-aminoacid isoform of amyloid beta, Aβ40 40-aminoacid isoform of amyloid beta, t-Tau total Tau protein, p-Tau hyperphosphorylated Tau protein

all comparisons), and also in the SNAP in comparison with the HL group (p < 0.001). Subjects in the HL group were also more often APOE-ε4 carriers (67%) than in all other groups (p < 0.001).

### Effect of including the Aβ42/40 ratio in MCI biomarkerbased classification

When we used the A $\beta$ 42/40 ratio, instead of A $\beta$ 42 alone, as a marker of amyloidosis, the MCI biomarker-based classification changed according to what is presented in Table 3. In MCI subtypes with already conclusive biomarker profiles (LL and HL), use of the A $\beta$ 42/40 ratio did not change their classification in 87% of cases (83% of LL and 90% of HL). Noteworthy, in 10/59 patients classified previously as LL, the Aβ42/40 ratio fell below the cutoff value and they were therefore classified as IAP; likewise, in 6/62 patients classified previously as HL, the A $\beta$ 42/40 ratio was within the normal range and changed their classification to SNAP. Regarding the subgroups with conflicting biomarker results, more pronounced changes were seen both in the IAP group (with 7/21 (33%) changing into the LL category) and particularly in the SNAP group (where 36/55 (66%) of patients had an abnormal A\u00e342/40 ratio and therefore changed their classification to HL). Overall, addition of the  $A\beta42/40$  ratio resulted in a redistribution of the MCI subtypes to 56 (28.4%) LL, 92 (46.7%) HL, 24 (12.2%) IAP and 25 (12.7%) SNAP (Table 3), therefore significantly improving the proportion of interpretable biological profiles from 61% to 75% (p = 0.001). Moreover, it significantly increased the percentage of MCI patients with amyloidosis from 42 to 59% (p < 0.001), therefore reaching a similar percentage to the patients with injury markers. Overall demographic, clinical and genetic differences between these new subgroups are similar to those presented in Table 2 (Additional file 2: Table S1). Interestingly, among patients classified as SNAP according to Aβ42 alone, those who changed their classification to HL according to the Aβ42/40 ratio had significantly lower Aβ42 than those who remained in the SNAP category (751 ± 148 vs 1088 ± 343 pg/ml, p < 0.001). A similar pattern was also seen in patients classified previously in the LL group who changed their classification to IAP in relation to those who

<sup>\*</sup>p < 0.05 vs controls

<sup>\*</sup>p < 0.01 vs controls

<sup>\*\*\*\*</sup>p < 0.001 vs controls

 $<sup>^{\</sup>gamma}p < 0.05$  vs AD-dementia

 $<sup>\</sup>gamma \gamma p < 0.01$  vs AD-dementia  $\gamma \gamma p < 0.001$  vs AD-dementia

<sup>§</sup>p < 0.05 vs MCI-St

p < 0.01 vs MCI-St

<sup>§§§</sup>p < 0.001 vs MCI-St

Table 2 Demographic, clinical, genetic and biomarker data of the MCI subgroups based on core CSF biomarkers

	Low-AD likelihood	High-AD likelihood	IAP	SNAP
N (%)	59 (29.9%)	62 (31.5%)	21 (10.6%)	55 (27.9%)
Gender (M/F)	17/42	26/36	8/13	17/38
Age (years)	62.7 ± 9.9	69.6 ± 7.7**	$66.3 \pm 9.7$	69.4 ± 8.9**
Age onset (years)	59.0 ± 10.3	66.4 ± 7.8**	63.1 ± 8.1	67.4 ± 8.1**
Education (years)	$6.5 \pm 3.9$	6.1 ± 4.1	$5.1 \pm 2.4$	$5.8 \pm 4.1$
MMSE	27.6 ± 2.6	24.3 ± 4.2***	25.1 ± 6.0	26.3 ± 3.2
MoCA	$20.5 \pm 5.0$	16.4 ± 5.5**	$16.3 \pm 6.9$	17.3 ± 4.9
ADAS-Cog	$8.5 \pm 4.6$	12.7 ± 4.3**	$13.3 \pm 7.5$	13.5 ± 7.5*
APOE-ε4 (%)	26%	64%***	24% <sup>YYY</sup>	39% <sup>YY</sup>
Aβ42 (pg/ml)	918 ± 243	405 ± 109***	403 ± 123***,§§§	867 ± 282 <sup>YYY</sup>
Aβ40 (pg/ml)	9608 ± 3219	10,945 ± 4191	8006 ± 3082 <sup>Y,§§§</sup>	14,247 ± 5219***, <sup>YYY</sup>
Aβ42/40 ratio	$0.105 \pm 0.040$	0.040 ± 0.017***	0.066 ± 0.024***	$0.061 \pm 0.037***,777$
t-Tau (pg/ml)	169 ± 42	545 ± 274***	159 ± 67 <sup>YYY,§§§</sup>	488 ± 250***
p-Tau (pg/ml)	30 ± 9	68 ± 28***	$28 \pm 7^{\gamma\gamma\gamma,\$\$\$}$	61 ± 27***
Follow-up time (years)	$4.0 \pm 3.3$	$4.0 \pm 2.5$	4.2 ± 4.1	$3.6 \pm 2.7$

Data expressed as mean ± standard deviation, except for APOE expressed as percentage of £4 carriers

MCI mild cognitive impairment, AD Alzheimer's disease, IAP isolated amyloid pathology, SNAP suspected non-Alzheimer pathology, M male, F female, MMSE Mini Mental State Examination, MoCA Montreal Cognitive Assessment, ADAS-Cog Alzheimer Disease Assessment Scale—Cognitive, APOE Apolipoprotein E, Aβ42 42-aminoacid isoform of amyloid beta, Aβ40 40-aminoacid isoform of amyloid beta, t-Tau total Tau protein, p-Tau hyperphosphorylated Tau protein MMSE and MoCA, higher scores correspond to better performance; ADAS-Cog, lower scores correspond to better performance

remained in this category (Aβ42 = 758 ± 114 pg/ml vs Aβ42 = 951 ± 250 pg/ml, p = 0.015). Conversely, for patients previously classified into the HL group, significantly decreased t-Tau and p-Tau levels were found in those who changed their classification to SNAP (t-Tau = 310 ± 76 pg/ml, p-Tau = 42 ± 7 pg/ml) in relation to those who remained in this category (t-Tau = 570 ± 276 pg/ml, p = 0.001; p-Tau = 71 ± 28 pg/ml, p = 0.001). No other significant changes were observed in any of the demographic and clinical parameters between patients who changed their biomarker-based classification and those who did not.

**Table 3** Distribution of  $A\beta42/40$  ratio and further classification of MCI biomarker-based subgroups

	Aβ42/40 ratio < 0.068	Aβ42/40 ratio ≥ 0.068
LL (n = 59)	10 (17%)—IAP	49 (83%)—LL
IAP $(n = 21)$	14 (67%)—IAP	7 (33%)—LL
SNAP ( $n = 55$ )	36 (66%)—HL	19 (34%)—SNAP
HL (n = 62)	56 (90%)—HL	6 (10%)—SNAP

Data expressed as n (percentage) of patients

Aβ42/40 42-aminoacid isoform of amyloid beta/40-aminoacid isoform of amyloid beta, LL Low-AD likelihood, IAP isolated amyloid pathology, SNAP suspected non-Alzheimer pathology, HL high-AD likelihood, AD Alzheimer's disease

### Longitudinal assessment of MCI patients

Of the 197 MCI patients enrolled, 36 had follow-up < 2 years, 13 were dropouts and four patients were excluded from the further analysis because, although their clinical presentation was amnestic MCI, they developed frontotemporal dementia. The remaining 144 subjects with follow-up ≥ 2 years (mean follow-up time 4.6 ± 2.9 years, range 2-15 years) comprise the longitudinal study group, which was further dichotomized into those who were cognitively stable in the last observation (74 (51%); MCI-St) and those who progressed to dementia due to AD (70 (49%); MCI-AD). Mean follow-up time was not different between patients who converted to AD (4.7 ± 3.3 years) and those who did not (4.5  $\pm$  2.5 years, p =0.990). Baseline demographic, clinical, genetic and biomarker characteristics of MCI-St and MCI-AD patients are presented in Table 1. No differences in gender distribution or years of education were seen between groups, but MCI-AD patients were older both at baseline observation and at disease onset than MCI-St patients. As expected, MCI-AD patients presented with a more severe cognitive impairment at baseline (significantly lower MMSE and MoCA and higher ADAS-Cog scores) than MCI-St patients, and were also more frequently APOE-ε4 carriers. Differences in baseline CSF Aβ42, t-

<sup>\*</sup>p < 0.05 vs low-AD likelihood

<sup>\*\*</sup>p < 0.005 vs low-AD likelihood

<sup>\*\*\*</sup>p < 0.001 vs low-AD likelihood

 $<sup>^{\</sup>gamma}p$  < 0.05 vs high-AD likelihood

 $<sup>^{\</sup>gamma\gamma}p$  < 0.01 vs high-AD likelihood  $^{\gamma\gamma\gamma}p$  < 0.001 vs high-AD likelihood

<sup>§§\$</sup> p < 0.001 vs SNAP

Tau and p-Tau, and in A $\beta$ 42/40, t-Tau/A $\beta$ 42 and A $\beta$ 42/p-Tau ratios, but not in A $\beta$ 40, were seen between the two MCI groups. A $\beta$ 42, A $\beta$ 42/40 and A $\beta$ 42/p-Tau ratios were lower and t-Tau, p-Tau and the t-Tau/A $\beta$ 42 ratio were higher in MCI-AD patients, and were similar to the values presented by AD patients. MCI-St patients had comparable t-Tau, p-Tau, and t-Tau/A $\beta$ 42 and A $\beta$ 42/p-Tau ratios to the control group, while A $\beta$ 42 and the A $\beta$ 42/40 ratio were slightly but significantly lower than the controls. MCI-St patients also had significantly increased levels of A $\beta$ 40 in relation to AD patients.

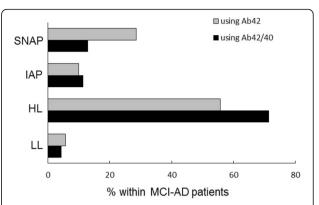
### Conversion to AD in MCI biomarker-based subgroups

Conversion to AD in MCI categories based on their core CSF biomarkers (Aβ42, t-Tau and p-Tau) showed that subjects in the HL group were more prone to progress to AD (75.0%), than all other biomarker groups: SNAP, 55.6%; IAP, 43.8%; LL, 10.0% (p < 0.001) (Table 4). This profile did not change much when we used the new classification with the Aβ42/40 ratio alongside Tau and p-Tau: progression to AD during clinical follow-up occurred in 72% of HL patients and in only 8% of the LL group. Noteworthy, still approximately half of the patients within the IAP (44%) and SNAP (53%) subtypes progressed to AD during clinical follow-up (Table 4). Interestingly, in patients classified previously into the IAP subgroup, conversion to AD occurred in 58.3% (7 out of 12) of patients who remained in this subgroup, and in none of the patients who changed their classification to LL, after replacing A $\beta$ 42 by the A $\beta$ 42/40 ratio. This difference, however, failed to reach statistical significance (p = 0.146). In patients classified previously into the SNAP subgroup, conversion to AD occurred at a similar percentage both in the patients who remained classified as SNAP after Aβ42/40 ratio inclusion (5 out of 11, 45.5%) and those who changed their classification to HL (15 out of 25, 60.0%) (p = 0.656). Figure 1 shows the percentage of the different MCI biomarker-based subgroups, determined using either the core CSF biomarkers (A $\beta$ 42, t-Tau and p-Tau) or the A $\beta$ 42/40 ratio,

**Table 4** Conversion to AD in the different MCI biomarker-based subgroups

	Using A $\beta$ 42, t-Tau and $p$ -Tau for classification	Using Aβ42/40 ratio, t-Tau and <i>p</i> -Tau for classification
LL	4/40 (10%)	3/38 (8%)
IAP	7/16 (44%)	8/18 (44%)
SNAP	20/36 (56%)	9/17 (53%)
HL	39/52 (75%)	50/71 (72%)

Data are expressed as n/total (percentage) of patients AD Alzheimer's disease, MCI mild cognitive impairment,  $A\beta42$  42-aminoacid isoform of amyloid beta, t-Tau total Tau protein, p-Tau hyperphosphorylated Tau protein,  $A\beta40$  40-aminoacid isoform of amyloid beta, LL low-AD likelihood, IAP isolated amyloid pathology, SNAP suspected non-Alzheimer pathology, HL High-AD likelihood



**Fig. 1** Comparison of MCI biomarker-based subgroups with clinical follow-up. Amongst mild cognitive impairment patients who progressed to Alzheimer's disease during clinical follow-up (MCI-AD), the percentage of different biomarker-based subgroups was compared. Biomarker-based subgroups determined using either CSF Aβ42, t-Tau and p-Tau (gray bars) or Aβ42/40 ratio, t-Tau and p-Tau (black bars). LL low-AD likelihood, HL high-AD likelihood, IAP isolated amyloid pathology, SNAP suspected non-Alzheimer pathology; Aβ42 42-aminoacid isoform of amyloid beta, Aβ40 40-aminoacid isoform of amyloid beta

t-Tau and p-Tau, amongst patients who progressed to AD during clinical follow-up. The percentage of LL and IAP subtypes within MCI-AD patients was low (6% and 11%, respectively) and was practically not influenced by the inclusion of A $\beta$ 42 (5.7% and 10.0%, respectively) or the A $\beta$ 42/40 ratio (4.3% and 11.4%, respectively) in their biomarker-based classification. On the contrary, the inclusion of the A $\beta$ 42/40 ratio significantly decreased the proportion of the SNAP subtype (from 28.6 to 12.9%) and increased the HL subtype (from 55.7 to 71.4%) within MCI-AD patients (p = 0.019).

### Predictors for AD-type dementia at follow-up

Logistic regression models were employed to identify the best predictors of conversion to AD. In the first model we included age, gender, education, follow-up time, MMSE, APOE genotype and CSF Aβ42, t-Tau and p-Tau values as variables in the equation and verified that the variables that were contributing significantly to the model classification were age (p = 0.012), CSF A $\beta$ 42 (p <0.001) and t-Tau (p = 0.033). We then substituted Aβ42 by the A $\beta$ 42/40 ratio in the model, and the variables retained in the model were again age (p = 0.015), CSF Aβ42/40 ratio (p < 0.001) and t-tau (p = 0.043). This last model that included the Aβ42/40 ratio showed a slightly better fit that the one without it, as the 2 log-likelihood or deviance (a measure for unexplained variance) was lower (91.5 vs 95.7, respectively). We then compared the ROC curves of the predicted probabilities derived from the two logistic regression models (Additional file 1: Figure S1). No statistically significant difference was seen between the AUC of the model including the

Aβ42/40 ratio (AUC = 0.898, 95% CI 0.839–0.956) or Aβ42 (AUC = 0.879, 95% CI 0.815–0.943) (p = 0.181).

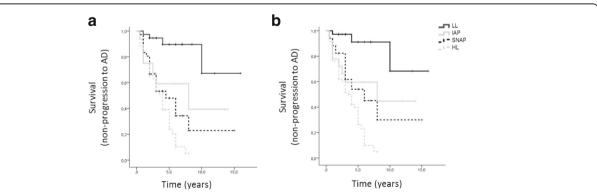
### Survival analysis of MCI biomarker-based subgroups

Figure 2 shows Kaplan-Meier survival curves for the probability of conversion to AD in the different MCI biomarker-based subgroups, determined using either Aβ42 or the Aβ42/40 ratio, in combination with Tau and p-Tau. When Aβ42 was used (Fig. 2a), the LL group was significantly associated with a longer estimated time of conversion to AD (13.3 ± 1.4 years, 95% CI 10.6–15.9) than all other groups (HL,  $3.6 \pm 0.3$  years, 95% CI 3.0-4.3, p < 0.001; SNAP, 6.3 ± 1.1 years, 95% CI 4.0–8.5, p <0.001; IAP, 7.7  $\pm$  1.7 years, 95% CI 4.5–11.0, p = 0.009). Estimated time to conversion was no different between the IAP and SNAP groups (p = 0.483), while a statistically significant difference was seen between the HL and IAP groups (p = 0.046), but not between HL and SNAP (p =0.095). In Cox regression models with age, gender, education, ApoE genotype and baseline MMSE score taken into account, the difference between the HL and IAP groups was no longer significant, and a quite similar increased risk of conversion was seen in the HL (hazard ratio 7.0, 95% CI 1.9–25.6, p = 0.003), SNAP (hazard ratio 5.9, 95% CI 1.6–20.9, p = 0.006) and IAP (hazard ratio 6.3, 95% CI 1.6-25.6, p = 0.009) subtypes compared with patients classified into the LL group (reference). When, instead of A $\beta$ 42, the A $\beta$ 42/40 ratio was considered (Fig. 2b), a statistically decreased estimated time of conversion was again seen in HL (3.7  $\pm$  0.3 years, 95% CI 3.1–4.2, p < 0.001), IAP (8.1  $\pm$  1.5 years, 95% CI 5.1-11.1, p = 0.006) and SNAP (7.3  $\pm$  1.6 years, 95% CI 4.3–2.4, p = 0.002) in comparison with the LL subgroup (13.5 ± 1.4 years, 95% CI 10.8-16.2). No difference was seen between the two conflicting subgroups (IAP and SNAP, p = 0.787), but estimated time to conversion was statistically lower in the HL group in relation both to IAP (p = 0.016) and SNAP (p =0.029). The Cox regression model also showed that MCI patients belonging to the HL subtype had the highest risk of progression to AD (hazard ratio 10.1, 95% CI 2.2-43.0, p = 0.003), compared with patients classified into the LL group (reference). MCI patients classified into the IAP and SNAP subtypes also presented an increased risk of progression to AD in comparison with the LL subtype (IAP, hazard ratio 8.0, 95% CI 1.7–41.0, p = 0.008; SNAP, hazard ratio 6.1, 95% CI 1.2-30.7, p = 0.029). However, risk of progression to AD failed to reach statistical significance between the HL group and the IAP or the SNAP group. Importantly, the hazard ratio of the HL group in this model with the Aβ42/40 ratio was considerably larger than that of the same group in the Cox regression model with Aβ42.

### **Discussion**

In this study we investigated the effect of using the CSF A $\beta$ 42/40 ratio, instead of A $\beta$ 42 levels, both combined with Tau and p-Tau, in MCI classification according to the NIA-AA criteria. Our main findings were that the percentage of patients with amyloidosis (HL + IAP) significantly increased from 42 to 59%, reaching a similar percentage to the patients with injury markers (HL + SNAP, 56%). The proportion of interpretable biological profiles also increased significantly from 61% to 75%, particularly due to a reduction to half in the number of SNAP cases.

In our work we found an overall increase in CSF A $\beta$ 40 in MCI patients in relation to AD-dementia patients, and no difference in relation to controls. A few authors have reported on the levels of CSF A $\beta$ 40 in MCI patients, with increased [71], decreased [11] or unchanged levels [31, 32, 72] in relation to controls being reported. These discrepancies might be explained in part



**Fig. 2** Kaplan–Meier survival curves for probability of conversion to AD according to different MCl biomarker-based subgroups. Number of individuals at risk at each time interval shown below the graphs. MCl subgroups determined taking into account CSF t-Tau and p-Tau levels and either Aβ42 (**a**) or the Aβ42/40 ratio (**b**). Log-rank (Mantel–Cox) p < 0.001 for both. AD Alzheimer's disease, LL low-AD likelihood, HL high-AD likelihood, IAP isolated amyloid pathology, SNAP suspected non-Alzheimer pathology

by technical differences in relation to the antibodies used in the different assays. While initial assays used N-terminally unspecific antibodies that could capture also Nterminally shortened A\u00e340 isoforms, this has changed, and Aβ40 assays like the one we used in this work are now based on N-terminally specific antibodies. In spite of this discrepancy related to CSF Aβ40 levels, most data, including ours, point toward a decreased CSF Aβ42/40 ratio in MCI patients in relation to controls. Reasonable agreement has also been reported regarding the cutoff value for the CSF Aβ42/40 ratio, ranging between 0.05 and 0.082 [11, 23, 27-29]. In this work, we established a cutoff value of 0.068 for distinguishing AD-dementia from controls, which is in line with the results of other groups regardless of using the same commercial assay or not. In fact, in the multicenter study by Dumurgier et al. [29] the mean Aβ42/40 ratio was comparable across centers, despite the significant inter-center differences in reported CSF Aβ40 and Aβ42 levels. Therefore, this ratio seems to be less sensitive to preanalytical and analytical sources of variability both intra-laboratories [73–75] and inter-laboratories [29].

One interesting finding of our study was that CSF Aβ40 values were significantly more dispersed in MCI patients with an ambiguous profile of core CSF biomarkers. In fact, CSF Aβ40 was significantly higher in the subgroup of patients with high t-Tau/p-Tau but normal Aβ42 (SNAP), while it was lower in patients with low Aβ42 but normal levels of t-Tau/p-Tau (IAP). A similar finding had also been reported by Sauvée et al. [27] in a mixed population of patients with dementia. This supports the idea that a large inter-individual variability in Aβ load can occur and that the normalization of CSF Aβ42 concentrations to Aβ40, rather than using absolute values of Aβ42, is a more accurate measure of amyloidosis. It would be interesting to see whether indeed, as reported recently by other groups [30, 76], the Aβ42/40 ratio correlated better with amyloid load data from PET imaging than Aβ42 alone. Unfortunately, amyloid-PET was not widely available for our MCI cohort. One other possible explanation for the large dispersion in CSF Aβ40 levels that should be considered is the preanalytical variability between samples. This is considered an important confounding source for core CSF biomarkers, and can also have an impact on Aβ40 measurements, as described by others [77–79].

The use of the CSF A $\beta$ 42/40 ratio, instead of A $\beta$ 42, had a significant effect in MCI patient biomarker-based categorization. Only minor changes were seen in the classification of patients who already had a concordant biomarker profile (all markers normal or all markers abnormal). On the contrary, use of the A $\beta$ 42/40 ratio changed the classification of 50 out of 76 patients with previous ambiguous results (65.8%), and who now fell

into conclusive categories. This resulted in a significant increase in the proportion of interpretable biological profiles from 61% to 75%. This is in accordance with previous results reporting that the added value of the Aβ42/40 ratio was particularly seen in patients with a discrepancy between CSF p-Tau and Aβ42, leading to a new and more informative biological conclusion [27, 29]. Overall, the most obvious effect of the use of the Aβ42/40 ratio in MCI patient classification was an increase in the percentage of patients classified into the HL group (from 31.5 to 46.5%), at the expense of a reduction in the SNAP group (from 27.9 to 12.7%). Therefore, a significant increase in the percentage of MCI patients with a positive marker for amyloidosis was also observed (from 42.1 to 58.9%), now reaching a similar percentage to the patients with injury markers. The fact that the normalization of Aβ42 using Aβ40 resulted in an approximately 50% reduction of the patients in the SNAP category is in favor of the hypothesis that a methodological bias (too conservative CSF Aβ42 cutoff values) may be underlying the unexpected high prevalence of the SNAP group reported previously [35, 37, 39]. Indeed, we observed that the mean CSF Aβ42 was lower in MCI patients who changed from SNAP to HL after taking into account the Aβ42/40 ratio than in those who remained classified as SNAP.

Amongst the 144 MCI patients who completed longitudinal clinical evaluation, no difference in baseline CSF Aβ40 was seen between MCI-St and MCI-AD patients, as already reported by others [31, 32], whereas the Aβ42/40 ratio was significantly lower in the MCI-AD group. Interestingly, our results show that both A\u00e342 and the A $\beta$ 42/40 ratio were statistically different (lower) between MCI-St patients and controls, while t-Tau and p-Tau levels were comparable between the two groups. This observation might indicate that some of the subjects who have not yet progressed to AD at the time of follow-up observation, and were therefore included in the MCI-St group, will indeed progress to AD in the future, as they have decreased Aβ42 and/or Aβ42/40 ratio levels. If we consider that amyloid alterations precede neurodegeneration in AD, we may assume that these subjects are at an early stage of the disease and would progress to AD dementia if observed for a longer time. In fact, this MCI-AD/MCI-St dichotomization is completely dependent on the follow-up time, with longer periods of observation giving rise to more accurate data.

In accordance with previous studies [35, 38–40, 42], the HL group showed the highest risk of progression to AD, irrespective of the use of either A $\beta$ 42 or the A $\beta$ 42/40 ratio for biomarker-based classification. Regarding prognosis in the conflicting biomarker categories, in our study the percentage of MCI patients classified as IAP or SNAP who converted to AD was not very different

from each other (close to 50%) and did not differ much whether CSF Aβ42 alone or the Aβ42/40 ratio, combined with Tau and p-Tau, was used. Nevertheless, the results depicted in Fig. 1 show that, within MCI patients who progressed to AD, the use of the Aβ42/40 ratio resulted in a significant increase in the percentage of patients classified as HL and a decrease in the percentage of patients classified as SNAP. Survival analysis showed that, when Aβ42 was used for MCI subgroup classification, both IAP and SNAP had significantly lower estimated time of conversion to AD than the LL group, that was similar to the HL group in the case of SNAP but not in the case of the IAP subgroup. The inclusion of the  $A\beta42/40$  ratio in MCI biomarker-based subgroup classification decreased the overlap between the HL and the SNAP survival curves, with both conflicting subgroups now showing an estimated time of conversion to AD that was significantly lower than the LL subgroup and higher than the HL subgroup. In Cox regression models, the same overall effect of the Aβ42/ 40 ratio on the risk of progression to AD was seen. In the model that used Aβ42, risk of progression to AD was very similar between the HL, IAP and SNAP subgroups. When we used the Aβ42/40 ratio instead, the difference between the hazard ratios of the three potential risk categories (IAP, SNAP and HL) increased. Both the IAP and SNAP categories presented with an equivalently increased risk of progression to AD, in comparison with the LL subtype, that was numerically lower than the HL group, but failed to reach statistical significance. More importantly, when comparing the hazard ratio of the HL group in both Cox regression models, it is evident that this is higher in the model that includes the  $A\beta42/40$  ratio than in the model that includes  $A\beta42$ (10.1 vs 7.0). This is also true for the IAP category (8.0 vs 6.3), but not for SNAP (6.1 vs 5.9). This finding is indicative that use of the Aβ42/40 ratio for patient classification indeed results in a better predictive value of future conversion to AD dementia. This was not confirmed by our logistic regression models, however, that failed to reach a statistically significant difference between the AUCs when including the Aβ42/40 ratio or Aβ42 in the models. Although some of our group analysis did not prove statistically that the A $\beta$ 42/40 ratio predicted conversion to AD with more accuracy compared to A $\beta$ 42, we believe the fact that it reduced conflicting biomarker results and produced biomarkerbased subgroups with more clear-cut differences in the risk of progression to AD is of value for individual MCI patient clinical follow-up.

Some limitations of the current study must be addressed. The fact that CSF A $\beta$ 40 assays (that only became available in our laboratory in 2016) were all performed within a short period of time in previously

stored samples, and not in a sequentially routine setting, as the other CSF biomarkers, might introduce some methodological bias to the results, and can contribute to the ratio having a better performance than Aβ42 alone. As already mentioned, imaging biomarkers were not considered in this study, as the availability of amyloid-PET data was very scarce. It would be interesting to correlate the CSF amyloid assessment either through Aβ42 or the Aβ42/40 ratio with this alternative diagnostic tool, categorizing the population as amyloid-positive or negative. Since only the amnestic subtype of MCI was considered, the generalization of the results to other forms of MCI should be cautious. Finally, as in many other clinical studies, no neuropathological verification was available, leaving the possibility of misdiagnosis. However, this study was developed in this specific context of routine clinical practice and we believe that this is a strength of the present work. Since we enrolled patients in a systematic way, our cohort may be considered representative of an ordinary tertiary Memory Clinic, surpassing the selection biases of investigational studies. Also, the fact that our data are based on a variable length of follow-up ( $\geq 2$  years) not only optimizes the available study information, but also reduced the chances of underestimating the predictive power of the selected parameters that might occur with a short fixed follow-up period. In addition, the rigorous methodology adopted to define stages and progression, the use of neuropsychological instruments well-validated for the Portuguese population and administered by the same experienced team of neuropsychologists, as well as the standardized use of the CSF biomarkers may also improve the reliability of the results.

### **Conclusion**

Our results confirm the usefulness of the addition of the CSF A $\beta$ 42/40 ratio in the interpretation of the CSF profile of MCI patients, as it increases the proportion of patients with conclusive profiles, therefore enhancing their predictive value for underlying AD dementia.

### Additional files

**Additional file 1: Figure S1.** Receiver operating characteristics (ROC) curves for (A) distinguishing between AD-dementia patients and controls and for (B) the predicted probabilities of conversion to AD in MCI patients derived from logistic regression models, using either CSF A $\beta$ 42 or the A $\beta$ 42/40 ratio (PDF 59 kb)

**Additional file 2: Table S1.** Demographic, clinical and genetic data of the MCI subgroups based on a CSF biomarker profile that included the  $A\beta42/40$  ratio, t-Tau and p-Ta. (PDF 216 kb)

### **Abbreviations**

AD: Alzheimer's disease; ADAS-Cog: Alzheimer Disease Assessment Scale—Cognitive; ANOVA: Analysis of variance; APOE: Apolipoprotein E; APP: Amyloid precursor protein; AUC: Area under the receiver operating

characteristics curve; AB: Amyloid beta; AB40: 40-aminoacid isoform of amyloid beta; Aβ42: 42-aminoacid isoform of amyloid beta; BLAD: Bateria de Lisboa para Avaliação de Demência (Lisbon Battery for Dementia Assessment); CDR: Clinical Dementia Rating scale; CI: Confidence interval; CNS: Central nervous system; CSF: Cerebrospinal fluid; CT: Computed tomography; CV: Coefficient of variance; DAD: Disability Assessment for Dementia; DSM-IV-TR: Diagnostic and Statistics Manual for Mental Disorders—fourth edition; EDTA: Ethylenediamine tetraacetic acid; ELISA: Enzyme-linked immunosorbent assay; HL: High-AD likelihood; IAP: Isolated amyloid pathology; LL: Low-AD likelihood; MCI: Mild cognitive impairment; MCI-AD: Mild cognitive impairment patients who progress to Alzheimer's disease; MCI-St: Stable mild cognitive impairment; MMSE: Mini Mental State Examination; MoCA: Montreal Cognitive Assessment; MRI: Magnetic resonance imaging; NIA-AA: National Institute of Aging-Alzheimer Association; NINCDS-ADRDA: National Institute of Neurological and Communicative Disorders and Stroke–Alzheimer's Disease and Related Disorders; NPI: Neuropsychiatric Inventory; PCR-RFLP: Polymerase chain reaction-restriction fragment length polymorphism; PET: Positron emission tomography; p-Tau: hyperphosphorylated Tau protein; ROC: Receiver operating characteristics; SD: Standard deviation; SNAP: Suspected non-Alzheimer pathology; SPECT: Single-photon emission computed tomography; SPSS: Statistical Package for the Social Sciences; STARD: Standards for Reporting of Diagnostic Accuracy; t-Tau: total Tau protein

### Acknowledgements

The authors wish to thank all patients and their caregivers who participated in the study.

### **Funding**

MJL was supported by the Portuguese Foundation for Science and Technology (reference PD/BD/135108/2017). DD was supported by the Portuguese Foundation for Science and Technology (ref SFRH/BD/52289/2013). The funding agency had no role in the study design, sample collection, data analysis or writing of the manuscript.

### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

### Authors' contributions

IB conceptualized the study, performed CSF biomarker analysis, analyzed the data and drafted the manuscript. IS made substantial contributions to study concept and design, coordinated clinical assessment and critically reviewed the manuscript. MJL and RP contributed to sample preparation and biomarker analysis. HG, MT-P and JB-C contributed to clinical assessment and follow-up, sample collection and database development. DD performed the neuropsychological evaluation and follow-up. MRA was responsible for genetic analysis. CRO contributed to study concept, supervised the research and critically revised the manuscript. All authors revised the manuscript for important intellectual content and gave final approval of the version to be published. All authors read and approved the final manuscript.

### Ethics approval and consent to participate

This study was performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and was approved by the Ethics Board of Coimbra University Hospital. All subjects or responsible caregivers, whichever appropriate, gave their informed consent.

### Consent for publication

Not applicable.

### **Competing interests**

The authors declare that they have no competing interests.

### **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

### **Author details**

<sup>1</sup>Laboratory of Neurochemistry, Neurology Department, Centro Hospitalar e Universitário de Coimbra, 3000-075 Coimbra, Portugal. <sup>2</sup>Center for

Neuroscience and Cell Biology, University of Coimbra, 3004-504 Coimbra, Portugal. <sup>3</sup>Faculty of Medicine, University of Coimbra, 3000-548 Coimbra, Portugal. <sup>4</sup>Dementia Clinic, Neurology Department, Centro Hospitalar e Universitário de Coimbra, 3000-075 Coimbra, Portugal. <sup>5</sup>Research & Development Unit, Centro Hospitalar e Universitário de Coimbra, 3000-075 Coimbra, Portugal.

## Received: 15 December 2017 Accepted: 25 February 2018 Published online: 20 March 2018

### References

- Prince M, Wimo A, Guerchet M, Ali G, Wu Y, Prina M. World Alzheimer Report 2015. The global impact of dementia. An analysis of prevalence, incidence, cost & trends. London: Alzheimer's Disease International (ADI): 2015.
- Morris JC. Early-stage and preclinical Alzheimer disease. Alzheimer Dis Assoc Disord. 2005;1:163–76.
- Jansen WJ, Ossenkoppele R, Knol DL, Tijms BM, Scheltens P, FRJ V, Visser PJ, Amyloid Biomarker Study Group. Prevalence of cerebral amyloid pathology in persons without dementia: a meta-analysis. JAMA. 2015;313:1924–38.
- Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. Lancet Neurol. 2010;9:119–28.
- McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR Jr, Kawas CH, Klunk WE, Koroshetz WJ, Manly JJ, Mayeux R, Mohs RC, Morris JC, Rossor MN, Scheltens P, Carrillo MC, Thies B, Weintraub S, Phelps CH. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011;7:263–9.
- Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, Gamst A, Holtzmn DM, Jaqust WJ, Petersen RC, Synder PJ, Carrillo MC, Thies B, Phelps CH. The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011;7:270–9.
- Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, Iwatsubo T, Jack CR Jr, Kaye J, Montine TJ, Park DC, Reiman EM, Rowe CC, Siemers E, Stern Y, Yaffe K, Carrillo MC, Thies B, Morrison-Bogorad M, Wagster MV, Phelps CH. Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011;7:280–92.
- Blennow K, Hampel H, Weiner M, Zetterberg H. Cerebrospinal fluid and blood biomarkers in Alzheimer's diseases. Nat Rev Neurol. 2010;6:131–44.
- Olsson B, Lautner R, Andreasson U, Öhrfelt A, Portelius E, Bjerke M, Hölttä M, Rosén C, Olsson C, Strobel G, Wu E, Dakin K, Petzold M, Blennow K, Zetterberg H. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. Lancet Neurol. 2016;15:673–84.
- Hansson O, Zetterberg H, Buchhave P, Londos E, Blennow K, Minthon L. Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: a follow-up study. Lancet Neurol. 2006;5:228–34.
- Hertze J, Minthon L, Zetterberg H, Vanmechelen E, Blennow K, Hansson O. Evaluation of CSF biomarkers as predictors of Alzheimer's disease: a clinical follow-up study of 4.7 years. J Alzheimers Dis. 2010;21:1119–28.
- Shaw L, Vanderstichele H, Knapik-Czajka M, Clark CM, Aisen PS, Petersen RC, Blennow K, Soares H, Simon A, Lewczuk P, Dean R, Siemers E, Potter W, Lee VM, Trojanowski JQ. Alzheimer's Disease Neuroimaging Initiative. Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. Ann Neurol. 2009;65:403–13.
- Mattsson N, Zetterberg H, Hansson O, Andreasen N, Parnetti L, Jonsson M, Herukka SK, van der Flier WM, Blankenstein MA, Ewers M, Rich K, Kaiser E, Verbeek M, Tsolaki M, Mulugeta E, Rosén E, Aarsland D, Visser PJ, Schröder J, Marcusson J, de Leon M, Hampel H, Scheltens P, Pirttilä T, Wallin A, Jönhagen ME, Minthon L, Winblad B, Blennow K. CSF biomarkers and incipient Alzheimer disease in patients with mild cognitive impairment. JAMA. 2009;302:385–93.
- 14. Visser PJ, Verhey F, Knol DL, Scheltens P, Wahlund LO, Freund-Levi Y, Tsolaki M, Minthon L, Wallin AK, Hampel H, Bürger K, Pirttila T, Soininen H, Rikkert

- MO, Verbeek MM, Spiru L, Blennow K. Prevalence and prognostic value of CSF markers of Alzheimer's disease pathology in patients with subjective cognitive impairment or mild cognitive impairment in the DESCRIPA study: a prospective cohort study. Lancet Neurol. 2009;8:619–27.
- Blennow K, Dubois B, Fagan AM, Lewczuk P, de Leon MJ, Hampel H. Clinical utility of cerebrospinal fluid biomarkers in the diagnosis of early Alzheimer's disease. Alzheimers Dement. 2015;11:58–69.
- Leitão MJ, Baldeiras I, Herukka SK, Pikkarainen M, Leinonen V, Simonsen AH, Perret-Liaudet A, Fourier A, Quadrio I, Veiga PM, de Oliveira CR. Chasing the effects of pre-analytical confounders—a multicenter study on CSF-AD biomarkers. Front Neurol. 2015;6:153.
- 17. Lelental N, Brandner S, Kofanova O, Blennow K, Zetterberg H, Andreasson U, Engelborghs S, Mroczko B, Gabryelewicz T, Teunissen C, Mollenhauer B, Parnetti L, Chiasserini D, Molinuevo JL, Perret-Liaudet A, Verbeek MM, Andreasen N, Brosseron F, Bahl JM, Herukka SK, Hausner L, Frölich L, Labonte A, Poirier J, Miller AM, Zilka N, Kovacech B, Urbani A, Suardi S, Oliveira C, Baldeiras I, Dubois B, Rot U, Lehmann S, Skinningsrud A, Betsou F, Wiltfang J, Gkatzima O, Winblad B, Buchfelder M, Kornhuber J, Lewczuk P. Comparison of different matrices as potential quality control samples for neurochemical dementia diagnostics. J Alzheimers Dis. 2016;52:51–64.
- Lewczuk P, Riederer P, O'Bryant SE, Verbeek MM, Dubois B, Visser PJ, Jellinger KJ, Engelborghs S, Ramirez A, Parnetti L, Jack Jr CR, Teunissen CE, Hampel H, Lleó A, Jessen F, Glodzik L, de Leon MJ, Fagan AM, Molinuevo JL, Jansen WJ, Winblad B, Shaw LM, Andreasson U, Otto M, Mollenhauer B, Wiltfang J, Turner MR, Zerr I, Handels R, Thompson AG, Johansson G, Ermann N, Trojanowski JQ, Karaca I, Wagner H, Oeckl P, van Waalwijk van Doorn L, Bjerke M, Kapogiannis D, Kuiperij HB, Farotti L, Li Y, Gordon BA, Epelbaum S, Vos SJB, Klijn CJM, Van Nostrand WE, Minguillon C, Schmitz M, Gallo C, Lopez Mato A, Thibaut F, Lista S, Alcolea D, Zetterberg D, Blennow K, Kornhuber J; Members of the WFSBP Task Force Working on this Topic: Riederer P, Gallo C, Kapogiannis D, Lopez Mato A, Thibaut F. Cerebrospinal fluid and blood biomarkers for neurodegenerative dementias: an update of the Consensus of the Task Force on Biological Markers in Psychiatry of the World Federation of Societies of Biological Psychiatry, World J Biol Psychiatry. 2017. https://doi.org/10.1080/15622975.2017.1375556.
- Portelius E, Westman-Brinkmalm A, Zetterberg H, Blennow K. Determination of beta-amyloid peptide signatures in cerebrospinal fluid using immunoprecipitation-mass spectrometry. J Proteome Res. 2006;5:1010–6.
- 20. LaFerla FM, Green KN, Oddo S. Intracellular amyloid-beta in Alzheimer's disease. Nat Rev Neurosci. 2007;8:499–509.
- Wiltfang J, Esselmann H, Bibl M, Hüll M, Hampel H, Kessler H, Frölich L, Schröder J, Peters O, Jessen F, Luckhaus C, Perneczky R, Jahn H, Fiszer M, Maler JM, Zimmermann R, Bruckmoser R, Kornhuber J, Lewczuk P. Amyloid beta peptide ratio 42/40 but not Aβ42 correlates with phospho-Tau in patients with low- and high-CSF Aβ40 load. J Neurochem. 2007;101:1053–9.
- Baldeiras I, Santana I, Leitão MJ, Ribeiro MH, Pascoal R, Duro D, Lemos R, Santiago B, Almeida MR, Oliveira CR. Cerebrospinal fluid Aβ40 is similarly reduced in patients with frontotemporal lobar degeneration and Alzheimer's disease. J Neurol Sci. 2015;358:308–16.
- Lewczuk P, Lelental N, Spitzer P, Maler JM, Kornhuber J. Amyloid-β 42/40 Cerebrospinal fluid concentration ratio in the diagnostics of Alzheimer's disease: validation of two novel assays. J Alzheimers Dis. 2015:43:183–91.
- Lewczuk P, Esselmann H, Otto M, Maler JM, Henkel AW, Henkel MK, Eikenberg O, Antz C, Krause W-R, do Reulbach U, Kornhuber J, Wiltfang J. Neurochemical diagnosis of Alzheimer's dementia by CSF Aβ42, Aβ42/Aβ40 ratio and total tau. Neurobiol Aging. 2004;25:273–81.
- Slaets S, Le Bastard N, Martin J-J, Sleegers K, Van Broeckhoven C, De Deyn PP, Engelborghs S. Cerebrospinal fluid Aβ1-40 improves differential dementia diagnosis in patients with intermediate P-tau181P levels. J Alzheimers Dis. 2013;36:759–67.
- Janelidze S, Zetterberg H, Mattsson N, Palmqvist S, Vanderstichele H, Lindberg O, van Westen D, Stomrud E, Minthon E, Blennow K, for the Swedish BioFINDER study group, Hansson O. CSF Aβ42/Aβ40 and Aβ42/ Aβ38 ratios: better diagnostic markers of Alzheimer disease. Ann Clin Transl Neurol. 2016;3:154–65.
- Sauvée M, DidierLaurent G, Latarche C, Escanyé MC, Olivier JL, Malaplate-Armand C. Additional use of Aβ42/Aβ40 ratio with cerebrospinal fluid

- biomarkers P-Tau and Ab42 increases the level of evidence of Alzheimer's disease pathophysiological process in routine practice. J Alzheimers Dis. 2014;41:377–86.
- Dorey A, Perret-Liaudet A, Tholance Y, Fourie A, Quadrio I. Cerebrospinal fluid Aβ40 improves the interpretation of Aβ42 concentration for diagnosing Alzheimer's disease. Front Neurol. 2015;6:247.
- Dumurgier J, Schraen S, Gabelle A, Vercruysse O, Bombois S, Laplanche JL, Peoc'h K, Sablonnière B, Kastanenka KV, Delaby C, Pasquier F, Touchon J, Hugon J, Paquet C, Lehmann S. Cerebrospinal fluid amyloid-β 42/40 ratio in clinical setting of memory centers: a multicentric study. Alzheimers Res Ther. 2015;7:30.
- Lewczuk P, Matzen A, Blennow K, Parnetti L, Molinuevo JL, Eusebi P, Kornhuber J, Morris JC, Fagan AM. Cerebrospinal fluid Aβ42/40 corresponds better than Aβ42 to amyloid PET in Alzheimer's disease. J Alzheimers Dis. 2017;55:813–22.
- Hansson O, Zetterberg H, Buchhave P, Andreasson U, Londos E, Minthon L, Blennow K. Prediction of Alzheimer's Disease Using the CSF Aβ42/Aβ40 ratio in patients with mild cognitive impairment. Dement Geriatr Cogn Disord. 2007;23:316–20.
- Parnetti L, Chiasserini D, Eusebi P, Giannandrea D, Bellomo G, De Carlo C, Padiglioni C, Mastrocola S, Lisetti V, Calabresi P. Performance of Aβ1-40, Aβ1-42, total Tau, and phosphorylated Tau as predictors of dementia in a cohort of patients with mild cognitive impairment. J Alzheimers Dis. 2012;29:229–38.
- Brys M, Pirraglia E, Rich K, Rolstad S, Mosconi L, Switalski R, Glodzik-Sobanska L, De Santi S, Zinkowski R, Mehta P, Pratico D, Saint Louis LA, Wallin A, Blennow K, de Leon MJ. Prediction and longitudinal study of CSF biomarkers in mild cognitive impairment. Neurobiol Aging. 2009;30:682–90.
- 34. Frölich L, Peters O, Lewczuk P, Gruber O, Teipel SJ, Gertz HJ, Jahn H, Jessen F, Kurz A, Luckhaus C, Hüll M, Pantel J, Reischies FM, Schröder J, Wagner M, Rienhoff O, Wolf S, Bauer C, Schuchhardt J, Heuser I, Rüther E, Henn F, Maier W, Wiltfang J, Kornhuber J. Incremental value of biomarker combinations to predict progression of mild cognitive impairment to Alzheimer's dementia. Alzheimers Res Ther. 2017;9:84.
- Petersen RC, Aisen P, Boeve BF, Geda YE, Ivnik RJ, Knopman DS, Mielke M, Pankratz VS, Roberts R, Rocca WA, Weigand S, Weiner M, Wiste H, Jack CR Jr. Mild cognitive impairment due to Alzheimer disease in the community. Ann Neurol. 2013;74:199–208.
- Guo LH, Alexopoulos P, Eisele T, Wagenpfeil S, Kurz A, Perneczky R. The National Institute on Aging-Alzheimer's Association research criteria for mild cognitive impairment due to Alzheimer's disease: predicting the outcome. Eur Arch Psychiatry Clin Neurosci. 2013;263:325–33.
- Duara R, Loewenstein DA, Shen Q, Barker W, Potter E, Varon D, Heurlin K, Vandenberghe R, Buckley C. Amyloid positron emission tomography with (18)F-flutemetamol and structural magnetic resonance imaging in the classification of mild cognitive impairment and Alzheimer's disease. Alzheimers Dement. 2013;9:295–301.
- Prestia A, Caroli A, van der Flier WM, Ossenkoppele R, Van Berckel B, Barkhof F, Teunissen CE, Wall AE, Carter SF, Schöll M, Choo IH, Nordberg A, Scheltens P, Frisoni GB. Prediction of dementia in MCI patients based on core diagnostic markers for Alzheimer disease. Neurology. 2013;80:1048–56.
- 39. Vos SJ, Verhey F, Frölich L, Kornhuber J, Wiltfang J, Maier W, Peters O, Rüther E, Nobili F, Morbelli S, Frisoni GB, Drzezga A, Didic M, van Berckel BN, Simmons A, Soininen H, Kloszewska I, Mecocci P, Tsolaki M, Vellas B, Lovestone S, Muscio C, Herukka SK, Salmon E, Bastin C, Wallin A, Nordlund A, de Mendonça A, Silva D, Santana I, Lemos R, Engelborghs S, Van der Mussele S, Alzheimer's Disease Neuroimaging Initiative, Freund-Levi Y, Wallin ÅK, Hampel H, van der Flier W, Scheltens P, Visser PJ. Prevalence and prognosis of Alzheimer's disease at the mild cognitive impairment stage. Brain. 2015;138:1327–38.
- Caroli A, Prestia A, Galluzzi S, Ferrari C, van der Flier WM, Ossenkoppele R, Van Berckel B, Barkhof F, Teunissen C, Wall AE, Carter SF, Schöll M, Choo IH, Grimmer T, Redolfi A, Nordberg A, Scheltens P, Drzezga A, Frisoni GB. Alzheimer's Disease Neuroimaging Initiative. Mild cognitive impairment with suspected nonamyloid pathology (SNAP): prediction of progression. Neurology. 2015;84:508–15.
- 41. Bertens D, Knol DL, Scheltens P, Visser PJ. Alzheimer's Disease Neuroimaging Initiative. Temporal evolution of biomarkers and cognitive markers in the asymptomatic, MCI, and dementia stage of Alzheimer's disease. Alzheimers Dement. 2015;11:511–22.

- Alexopoulos P, Werle L, Roesler J, Thierjung N, Gleixner LS, Yakushev I, Laskaris N, Wagenpfeil S, Gourzis P, Kurz A, Perneczky R, Alzheimer's Disease Neuroimaging Initiative (ADNI). Conflicting cerebrospinal fluid biomarkers and progression to dementia due to Alzheimer's disease. Alzheimers Res Ther. 2016:8:51.
- Baldeiras I, Santana I, Proenca MT, Garrucho MH, Pascoal R, Rodrigues A, Duro D, Oliveira CR. Oxidative damage and progression to Alzheimer's disease in patients with mild cognitive impairment. J Alzheimers Dis. 2010; 21:1165–77.
- Folstein MF, Folstein SE, McHugh PR. Mini-Mental State: a practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res. 1975;12:189–98.
- Guerreiro M, Silva AP, Botelho MA. Avaliação Breve do Estado Mental: Escalas e Testes na Demência. Lisbon: Grupo de Estudos de Envelhecimento Cerebral e Demência; 2003. p. 27–32.
- Nasreddine Z, Phillips NA, Bédirian V, Charbonneau S, Whitehead V, Collin I, Cummings JL, Chertkow H. The Montreal Cognitive Assessment, MoCA: a brief screening tool for mild cognitive impairment. J Am Geriatr Soc. 2005; 53:605–0
- Simões MR, Freitas S, Santana I, Firmino H, Martins C, Nasreddine Z, Vilar M. Montreal Cognitive Assessment (MoCA): Versão final portuguesa. Coimbra: Serviço de Avaliação Psicológica, Faculdade de Psicologia e de Ciências da Educação da Universidade de Coimbra; 2008.
- Mohs RC, Rosen WG, Davis KL. The Alzheimer's Disease Assessment Scale: an instrument for assessing treatment efficacy. Psychopharmacol Bull. 1983; 19:448–50.
- Rosen WG, Mohs RC, Davis KL. A new rating scale for Alzheimer's Disease. Am J Psychiatry. 1984;141:1356–64.
- Guerreiro M, Fonseca S, Barreto J. Escalas e Testes na Demência. Lisbon: Grupo de Estudos de Envelhecimento Cerebral e Demência; 2003. p. 33–49.
- 51. Guerreiro M. Contribution of neuropsychology to the study of dementias. PhD thesis. Lisbon: Faculty of Medicine of Lisbon; 1988.
- PhD thesis. Lisbon: Faculty of Medicine of Lisbon; 1988.

  52. Berg L. Clinical dementia rating (CDR). Psychopharmacol Bull. 1988;24:637–9.
- Gelinas I, Gauthier L, McIntyre M, Gauthier S. Development of a functional measure for persons with Alzheimer's disease: the Disability Assessment for Dementia Scale. Am J Occup Ther. 1999;53:471–81.
- Leitão O. Escalas e Testes na Demência. Lisbon: Grupo de Estudos de Envelhecimento Cerebral e Demência; 2008. p. 107–110.
- Cummings JL. The Neuropsychiatric Inventory: assessing psychopathology in dementia patients. Neurology. 1997;48:10–6.
- 56. Leitão O, Nina A. Escalas e Testes na Demência. Lisbon: Grupo de Estudos de Envelhecimento Cerebral e Demência; 2003. p. 67–86.
- Petersen RC, Stevens JC, Ganguli M, Tangalos EG, Cummings JL, DeKosky ST. Practice parameter: early detection of dementia: mild cognitive impairment (an evidence-based review). Report of the Quality Standards Subcommittee of the American Academy of Neurology. Neurology. 2001;56:1132–42.
- Freitas S, Simões MR, Alves L, Santana I. Montreal Cognitive Assessment (MoCA): normative study for the Portuguese population. J Clin Exp Neuropsychol. 2011;33:989–96.
- Roman GC, Tatemichi TK, Erkinjuntti T, Cummings JL, Masdeu JC, Garcia JH, Amaducci L, Orgogozo JM, Brun A, Hofman A, Moody DM, O'Brien MD, Yamaguchi T, Grafman J, Drayer BP, Bennett DA, Ficher M, Ogata J, Kokmen E, Bermejo F, Wolf PA, Gorelick PB, Bick KL, Pajeau AK, Bell MA, DeCarli C, Culebras A, Korczyn D, Bogousslavsky J, Hartmann A, Scheinberg P. Vascular dementia: diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. Neurology. 1993;43: 250–60.
- Jack CR Jr, Knopman DS, Weigand SD, Wiste HJ, Vemuri P, Lowe V, Kantarci K, Gunter JL, Senjem ML, Ivnik RJ, Roberts RO, Rocca WA, Boeve BF, Petersen RC. An operational approach to National Institute on Aging-Alzheimer's Association criteria for preclinical Alzheimer disease. Ann Neurol. 2012;71:765–75.
- American Psychiatric Association. Diagnostic and statistical manual of mental disorders. 4th ed. Washington, DC: American Psychiatric Association; 1994.
- 62. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's disease. Neurology. 1984;4:939–44.

- 63. del Campo M, Mollenhauer B, Bertolotto A, Engelborghs S, Hampel H, Simonsen AH, Kapaki E, Kruse N, Le Bastard N, Lehmann S, Molinuevo JL, Parnetti L, Perret-Liaudet A, Sáez-Valero J, Saka E, Urbani A, Vanmechelen E, Verbeek M, Visser PJ, Teunissen C. Recommendations to standardize preanalytical confounding factors in Alzheimer's and Parkinson's disease cerebrospinal fluid biomarkers: an update. Biomark Med. 2012;6:419–30.
- Baldeiras I, Santana I, Garrucho MH, Pascoal R, Lemos R, Santiago B, Oliveira CR. CSF biomarkers for the early diagnosis of Alzheimer's disease in a routine clinical setting—the first Portuguese study. Dermatol Sin. 2012;12:14–22.
- van Waalwijk van Doorn LJC, Kulic L, Koel-Simmelink MJA, Kuiperij HB, Versleijen AAM, Struyfs H, Twaalfhoven HAM, Fourier A, Engelborghs S, Perret-Liaudet A, Lehmann S, Verbeek MM, Vanmechelen EJM, Teunissen CE. Multicenter analytical validation of Aβ40 immunoassays. Front Neurol. 2017;8:310
  - Mattsson N, Andreasson U, Persson S, Arai H, Batish SD, Bernardini S, Bocchio-Chiavetto L, Blankenstein MA, Carrillo MC, Chalbot S, Coart E, Chiasserini D, Cutler N, Dahlfors G, Duller S, Fagan AM, Forlenza O, Frisoni GB, Galasko D, Galimberti D, Hampel H, Handberg A, Heneka MT, Herskovits AZ, Herukka SK, Holtzman DM, Humpel C, Hyman BT, Iqbal K, Jucker M, Kaeser SA, Kaiser E, Kapaki E, Kidd D, Klivenyi P, Knudsen CS, Kummer MP, Lui J, Lladó A, Lewczuk P, Li QX, Martins R, Masters C, McAuliffe J, Mercken M, Moghekar A, Molinuevo JL, Montine TJ, Nowatzke W, O'Brien R, Otto M, Paraskevas GP, Parnetti L, Petersen RC, Prvulovic D, de Reus HP, Rissman RA, Scarpini E, Stefani A, Soininen H, Schröder J, Shaw LM, Skinningsrud A, Skrogstad B, Spreer A, Talib L, Teunissen C, Trojanowski JQ, Tumani H, Umek RM, Van Broeck B, Vanderstichele H, Vecsei L, Verbeek MM, Windisch M, Zhang J, Zetterberg H, Blennow K. The Alzheimer's Association external quality control program for cerebrospinal fluid biomarkers. Alzheimers Dement. 2011;7:386-95.
- Crook R, Hardy J, Duff K. Single-day apolipoprotein E genotyping. J Neurosci Methods. 1994;53:125–7.
- Hanley JA, McNeil BJ. A method of comparing the areas under receiver operating characteristic curves. Radiology. 1983;148:839–43.
- Gabriel AJ, Almeida MR, Ribeiro MH, Durães J, Tábuas-Pereira M, Pinheiro AC, Pascoal R, Santana I, Baldeiras I. Association between butyrylcholinesterase and cerebrospinal fluid biomarkers in Alzheimer's disease patients. Neurosci Lett. 2017;641:101–6.
- Bossuyt PM, Reitsma JB, Bruns DE, Gatsonis CA, Glasziou PP, Irwig LM, Moher D, Rennie D, de Vet HC, Lijmer JG. The STARD statement for reporting studies of diagnostic accuracy: explanation and elaboration. Clin Chem. 2003:49:718.
- Struyfs H, Van Broeck B, Timmers M, Fransen E, Sleegers K, Van Broeckhoven C, De Deyn PP, Streffer JR, Mercken M, Engelborghs S. Diagnostic accuracy of cerebrospinal fluid amyloid-β isoforms for early and differential dementia diagnosis. J Alzheimers Dis. 2015;45:813–22.
- Kester MI, Scheffer PG, Koel-Simmelink MJ, Twaalfhoven H, Verwey NA, Veerhuis R, Twisk JW, Bouwman FH, Blankenstein MA, Scheltens P, Teunissen C, van der Flier WM. Serial CSF sampling in Alzheimer's disease: specific versus non-specific markers. Neurobiol Aging. 2012; 33:1591–8.
- Gervaise-Henry C, Watfa G, Albuisson E, Kolodziej A, Dousset B, Olivier JL, Jonveaux TR, Malaplate-Armand C. Cerebrospinal fluid Aβ42/Aβ40 as a means to limiting tube- and storage-dependent pre-analytical variability in clinical setting. J Alzheimers Dis. 2017;57:437–55.
- Lewczuk P, Beck G, Esselmann H, Bruckmoser R, Zimmermann R, Fiszer M, Bibl M, Maler JM, Kornhuber J, Wiltfang J. Effect of sample collection tubes on cerebrospinal fluid concentrations of tau proteins and amyloid beta peptides. Clin Chem. 2006;52:332–4.
- Slemmon JR, Shapiro A, Mercken M, Streffer J, Romano G, Andreasen N, Zetterberg H, Blennow K. Impact of cerebrospinal fluid matrix on the detection of Alzheimer's disease with Aβ42 and influence of disease on the total- Aβ42/Aβ40 ratio. J Neurochem. 2015;135:1049–58.
- Janelidze S, Pannee J, Mikulskis A, Chiao P, Zetterberg H, Blennow K, Hansson O. Concordance between different amyloid immunoassays and visual amyloid positron emission tomographic assessment. JAMA Neurol. 2017; https://doi.org/10.1001/jamaneurol.2017.2814.
- 77. Zimmermann R, Lelental N, Ganslandt O, Maler JM, Kornhuber J, Lewczuk P. Preanalytical sample handling and sample stability testing

- for the neurochemical dementia diagnostics. J Alzheimers Dis. 2011;25:739–45.
- Schipke CG, Jessen F, Teipel S, Luckhaus C, Wiltfang J, Esselmann H, Frölich L, Maier W, Rüther E, Heppner FL, Prokop S, Heuser I, Peters O. Long-term stability of Alzheimer's disease biomarker proteins in cerebrospinal fluid. J Alzheimers Dis. 2011;26:255–62.
- 79. Willemse E, van Uffelen K, Brix B, Engelborghs S, Vanderstichele H, Teunissen C. How to handle adsorption of cerebrospinal fluid amyloid  $\beta$  (1-42) in laboratory practice? Identifying problematic handlings and resolving the issue by use of the  $A\beta42/A\beta40$  ratio. Alzheimers Dement. 2017;13:885–92.

# Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit

